



Structure—Activity Study of L-Cysteine-Based N-Type Calcium Channel Blockers: Optimization of N- and C-Terminal Substituents

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Abstract—Synthesis and structure–activity relationship (SAR) studies of L-cysteine-based N-type calcium channel blockers are described. In the course of exploring SAR of the N- and C-terminal substituents, the L-cysteine derivative **4b** was found to be a potent N-type calcium channel blocker with an IC $_{50}$ value of 0.14 μ M on IMR-32 assay. Compound **4b** showed 12-fold selectivity for N-type over L-type calcium channels on AtT-20 assay. © 2002 Elsevier Science Ltd. All rights reserved.

Introduction

Voltage-sensitive calcium channels play crucial roles in various biological processes, and are classified into several subtypes on the basis of pharmacological and electrophysiological properties as L-, N-, P-, Q-, R- or T-type calcium channels. Among these calcium channels, the N-, P-, Q-, and R-type channels have all been shown to play key roles in neurotransmitter release. N-Type calcium channels are located at presynaptic terminals throughout neurons and directly mediate spinal transmission of pain signals from the peripheral to the central nervous system.

ω-Conotoxin MVIIA, a 25-amino acid peptide, is a selective blocker of N-type calcium channels that shows analgesic activity when administered intrathecally.² Over the last decade, synthetic efforts have focused on small-molecule, non-peptide N-type calcium channel blockers for analgesia or neuroprotection, since clinical observations were reported for ω-Conotoxin MVIIA.² A number of small-molecule blockers of N-type calcium channels have been reported,^{3,4} some of which have been shown to be active in analgesic models.⁴ However,

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most of these compounds also block the other calcium channel subtypes, including L-type channels. Although blocking of neuronal L-type calcium channels shows neuroprotective effects, inhibition of cardiac L-type calcium channels probably causes hypotensive side effects. Previously, we reported the discovery of a novel series of N-type calcium channel blockers. N-(t-Butoxycarbonyl)-L-cysteine derivative **2** was identified as a lead compound for L-cysteine-based N-type calcium channel blockers, which inhibited calcium influx into IMR-32 human neuroblastoma cells with an IC of 0.61 μ M. Here, we report the synthesis and in vitro structureactivity relationship (SAR) studies of modification of N- and C-terminal substituents of L-cysteine.

The compounds synthesized were evaluated for inhibitory activity against both N-type (IMR-32 assay^{7,9}) and L-type (AtT-20 assay^{8,9}) calcium channels, focusing on selectivity to reduce cardiovascular side effects due to blocking of L-type calcium channels (Fig. 1).

Chemistry

The synthesis of L-cysteine-based N-type calcium channel blockers is outlined as shown in Scheme 1. Treatment of L-cysteine with cyclohexylmethyl bromide in the presence of aqueous sodium hydroxide in ethanol

afforded sulfide **5**. Subsequent reaction of compound **5** with di-*tert*-butyl dicarbonate gave *N*-Boc-protected compound **6** from L-cysteine by one-pot procedure. Reaction of carboxylic acid **6** with appropriately substituted benzylamines in the presence of EDC and HOBt and subsequent deprotection gave intermediates **7** for N-terminal modification. Treatment of compounds **7** with *N*-Boc-(*R*)-thiazolidine-4-carboxylic acid in the presence of EDC and HOBt afforded compounds **4**. Compounds shown in Tables 1 and 2 were synthesized by the same method as described for compounds **4**. This procedure can be easily applied to solution-phase combinatorial synthesis using polymer-supported condensation reagents.

Compounds shown in Table 3 were synthesized from compound **4b** by deprotection followed by condensation with carboxylic acids or reductive amination with aldehydes.

Results and Discussion

The starting point in this study was an aspartic acidderived compound 1, which was identified as an initial

Figure 1.

Scheme 1. Reagents: (a) cyclohexylmethyl bromide, 2N NaOH, EtOH; (b) Boc₂O; (c) 4-methoxybenzylamine or 4-phenoxybenzylamine, EDC, HOBt, dichloromethane; (d) 4N HCl/dioxane; (e) *N*-Boc-(*R*)-thiazolidine-4-carboxylic acid, EDC, HOBt, dichloromethane.

lead for a novel series of N-type calcium channel blockers. Compound 1 inhibited calcium influx into IMR-32 cells with an IC $_{50}$ of 3.4 μM . In the process of exploring the SARs of this series of N-type calcium channel blockers using L-amino acids as structural motifs, compounds 2 and 3 were discovered as lead compounds for further modification. These two compounds have L-cysteine residues and showed improved in vitro N-type calcium channel blocking potency (IC $_{50}$ of 0.61 and 0.64 μM , respectively) compared to the initial lead compound 1. In this study, we investigated the SARs and further optimization of substituents at N- and C-termini using L-cysteine as a central scaffold.

The SAR of L-cysteine-based N-type calcium channel blockers was initially investigated for N-terminal substituent R_1 (Table 1). While replacement of the *tert*-butoxycarbonyl group with an *iso*-butoxycarbonyl group resulted in slight loss of activity (9, IC₅₀ 0.72 μ M), removal of the *tert*-butoxycarbonyl group resulted in 10-fold loss of inhibitory activity for N-type calcium

Table 1. Modification of the N-terminal acyl group R_1 of L-cysteine: in vitro inhibition of calcium influx in IMR-32 and AtT-20 assays

Compd	R ₁	$IC_{50} (\mu M)^a$	
		N-type (IMR-32)	L-type (AtT-20)
2	\downarrow_{\circ}	0.61	1.7
7a	Н	5.8	_
9	Y.	0.72	1.5
10		1.2	2.7
11	Cy.	0.92	2.3
12		1.3	1.9
4 a	Boc O	0.39	0.68
13	Boc O	0.88	1.9
8a	HZ S	2.7	3.5
14	N O	1.9	2.7

^aValues represent means of multiple determinations performed in duplicate.

channels in IMR-32 assay (7a, IC₅₀ 5.8 μ M). This observation suggested that the lipophilic substituents enhance potency; however, substitution of the carbamate moiety with an acyl group showed decreased potency by 2-fold (10, IC₅₀ 1.2 μ M) compared to compound 2a. Similarly, substitution with cycloalkyl group (11 and 12) also reduced N-type blocking activity. Screening of N-acyl substituents indicated that N-tertbutoxycarbonyl-thiazolidine-4-carbonyl increased Ntype inhibitory potency (4a, IC₅₀ 0.39 µM). Interestingly, compound 13, which has an L-proline residue instead of a thiazolidine-4-carbonyl group on the Nterminal, showed lower potency (IC₅₀ 0.88 μM). Comparison of 4a and 13 showed that the thiazolidine ring led to a further enhancement of potency. Further modification of the sulfur-containing ring structure was considered, but compound 8a (IC₅₀ 2.7 µM), which did not have a t-butoxycarbonyl group on the thiazolidine ring, and compound 14, which had thiazole-4-carbonyl moiety, showed very weak inhibitory activity for N-type calcium channels.

The inhibitory activity of compound **4a** for N-type calcium channels was also confirmed by electrophysiological study using IMR-32 cells (48% inhibition at $10 \mu M$, n=3). Therefore, the following SAR study was performed with compound **4a**, which has an *N-t*-butoxycarbonyl-thiazolidine-4-carbonyl at its N-terminus.

Table 2. Modification of the C-terminal amide group R₂ of L-cysteine: in vitro inhibition of calcium influx in IMR-32 and AtT-20 assays

Compd	R_2	IC ₅₀ (μM) ^a	
		N-type (IMR-32)	L-type (AtT-20)
4a	OMe	0.39	0.68
15	NMe ₂	0.61	0.95
16	NO ₂	0.60	0.50
17	ОН	1.8	2.8
18	OBzl	0.50	1.5
4b	OPh	0.14	1.7

^aValues represent means of multiple determinations performed in duplicate.

Modification of C-terminal substituent R_2 was next investigated (Table 2). Replacement of substituents on the benzene ring did not significantly affect the potency for compounds 15 ($R_2 = NMe_2$; IC_{50} 0.61 μ M) and 16 ($R_2 = NO_2$; IC_{50} 0.60 μ M). Although both of these compounds showed weaker activity than compound 4a, there was no difference in the N-type inhibitory activity between compounds 15 and 16. These results suggested that there was no influence of the electronic effect of the substituent on the benzene ring. On the other hand, compound 17 with a free phenol moiety showed 5-fold lower potency ($R_2 = OH$; IC_{50} 1.8 μ M). Compound 18, which had benzyloxy moiety, was synthesized to estimate steric effects. Compound 18 showed slightly lower activity than compound 4a.

As a consequence of modification of substituents on the benzene ring, compound **4b**, which had a phenoxy residue, was found to be the most potent N-type blocker among this series of compounds. Compound **4b** blocked N-type calcium channels with an IC₅₀ of 0.14 μ M and showed good selectivity over L-type calcium channels (selectivity ratio IC₅₀ L-type/N-type = 12).

Finally, the SAR of modification of the substituents on the nitrogen atom of thiazolidine was investigated (Table 3). In contrast to the results with modification of

Table 3. Modification of substituents R_3 on the nitrogen atom of the thiazolidine ring: in vitro inhibition of calcium influx in IMR-32 and AtT-20 assays

Compd	R_3	IC ₅₀ (μM) ^a	
		N-type (IMR-32)	L-type (AtT-20)
4b	\downarrow	0.14	1.7
8b	Н	0.29	2.0
19		0.12	0.87
20		0.36	1.2
21		0.33	0.44
22	MeO	0.12	0.93
23	MeO	0.20	0.60
24	HO	0.35	1.1

^aValues represent means of multiple determinations performed in duplicate.

the N-terminal acyl group R_1 (Table 1), no significant influence of substituents was observed on the N-type inhibitory activity. However, it should be noted that most of these compounds showed increased inhibitory activity and selectivity for N-type calcium channels compared to the compounds shown in Table 1. These results indicated that the phenoxy moiety played a crucial role in inhibitory potency and selectivity. Among the compounds shown in Table 3, compounds 19 (R_3 = methoxycarbonyl; IC_{50} 0.12 μ M) and 22 (R_3 =2-methoxyacetyl; IC_{50} 0.12 μ M) were equipotent to compound 4b.

In conclusion, the SAR study of a series of L-cysteine-based compounds led to the discovery of novel neuronal N-type calcium channel blockers. Compound 4b, which had phenoxy residue on the C-terminal benzene ring, was a potent N-type calcium channel blocker and was 12-fold more selective for N-type over L-type calcium channels

References and Notes

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- 7. IMR-32 cells were grown as described (Clementi, F.; Cabrini, D.; Gotti, C.; Sher, E. *J. Neurochem.* **1986**, 47, 291. Carbone, E.; Sher, E.; Clementi, F. *Pflügers Arch.* **1990**, 416, 170) in Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal bovine serum (FBS), 100 IU/mL penicillin and 100 µg/mL streptomycin.
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- 9. $[Ca^{2+}]_I$ was measured in cell suspension. Cell suspensions were incubated with 5 mM fura-2/AM for 30 min at 37 °C. Cells were resuspended in Krebs–Ringer HEPES solution and adjusted to 1.0×10^6 cells/mL (IMR-32), or 2.0×10^6 cells/mL (AtT20/D16v-F2). Fluorescence (λ_{Ex} : 340 and 380 nm; λ_{Em} : 500 nm) was detected with a fluorometer. Cell suspensions were incubated with test compound and 10 μ M nifedipine (IMR-32) or 3 μ M ω -conotoxin MVIIC (AtT-20/D16v-F2) for 360 s before high-K $^+$ stimulus. To evaluate the inhibitory activities of test compounds, IMR-32 and AtT-20/D16v-F2 cells were used for N-type and L-type calcium channels, respectively.
- 10. The electrophysiological recordings were performed in the conventional whole-cell configuration under voltage-clamp conditions. Pipettes had a resistance of 3–6 $M\Omega$. Membrane currents were measured using a patch-clamp amplifier (Axopatch 2B Axon Instruments). The test compounds were applied using a rapid application method designated as the 'Y-tube method'.